### ANNUAL PROGRESS REPORT

#### FISH RESEARCH PROJECT OREGON

PROJECT TITLE: Spring Chinook Salmon in the Willamette and Sandy Rivers

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# **CONTENTS**

# **KEY FINDINGS**

- <span id="page-2-0"></span>1. The number of wild spring chinook salmon in 2002 in the four rivers where we can estimate adult runs from otolith analysis was:
	- 3,428 (McKenzie above Leaburg Dam),
	- 536 (North Santiam above Bennett Dam),
	- 1,493 (Clackamas above North Fork Dam), and
	- 954 (Sandy above Marmot Dam).
- 2. The percentage of wild spring chinook incorporated into hatchery broodstocks in 2002 was:
	- $\blacksquare$  1.2% (McKenzie),
	- 0.6% (North Santiam),
	- 2.1% (South Santiam), and
	- 0.3% (Willamette).

The general guideline in draft Hatchery Genetic Management Plans (HGMP) is 10%, although the HGMP for Willamette Hatchery identifies wild spring chinook as probably extinct in the Middle Fork Willamette.

- 3. We recovered a high number of wild spring chinook carcasses in 2002 in the South Santiam River (224), second only to the McKenzie (454).
- 4. Age 0 (subyearling) chinook salmon were found throughout the lower McKenzie, upper and lower Willamette, and Santiam rivers in late May–July, 2002 and 2003.
- 5. Using DNA microsatellite analysis, we found that 94% of the subyearling chinook salmon seined in 2002 in the Willamette River downstream of the Santiam River were spring chinook and 6% were introduced fall chinook. The racial composition did not change between June and July. The percentage of spring chinook in the lower Santiam River was 66%.
- 6. We documented that some subyearling chinook salmon (including some spring chinook) migrated past Willamette Falls in late spring to early summer. These fish were PIT-tagged in the Willamette, lower McKenzie, and Santiam rivers. Migration was higher for fish tagged in the lower Willamette and Santiam rivers.

#### **INTRODUCTION**

<span id="page-3-0"></span>The Willamette and Sandy rivers support intense recreational fisheries for spring chinook salmon (*Oncorhynchus tshawytscha*). Fisheries in these basins rely primarily on annual hatchery releases of 5–8 million juveniles. Hatchery programs exist in the McKenzie, Middle Fork Willamette, North and South Santiam, Clackamas, and Sandy rivers mainly as mitigation for dams that blocked natural production areas. Some natural spawning occurs in most of the major basins and a few smaller tributaries upstream of Willamette Falls.

The Oregon Fish and Wildlife Commission adopted the Native Fish Conservation Policy (ODFW 2003a) and the Hatchery Management Policy (ODFW 2003b) in part to reduce adverse impacts of hatchery programs on wild native stocks. The Native Fish Conservation Policy recognizes that naturally produced native fish are the foundation for long-term sustainability of native species and hatchery programs, and the fisheries they support.

In the past, hatchery programs and fish passage issues were the focus of spring chinook salmon management in the Willamette and Sandy basins. Limited information was collected on the genetic structure among basin populations, on abundance and distribution of natural spawning, on rearing and migrating of juvenile salmon, or on strategies for reducing risks that large hatchery programs pose for wild salmon populations. This study is being implemented to gather this information. A schematic of the study plan is shown in **APPENDIX A**.

We conducted work in the main-stem Willamette River at Willamette Falls, and in the Middle Fork Willamette, McKenzie, Calapooia, North Santiam, South Santiam, Molalla, Clackamas, and Sandy rivers in 2003. Basin descriptions and background information on management and fish runs can be found in subbasin plans developed by the Oregon Department of Fish and Wildlife (ODFW 1988, ODFW 1992a, ODFW 1992b, and ODFW 1996). Task headings below cross reference the study plan outlined in **APPENDIX A.** This report covers tasks that were worked on in late 2002 through early fall 2003.

#### **TASK 1.2–THE PROPORTION OF WILD FISH IN NATURAL SPAWNING POPULATIONS**

Implementation of the Native Fish Conservation Policy (and the Wild Fish Management Policy that preceded it) requires information on hatchery and wild fish in spawning populations. In response to this need and to implement a selective fishery, all hatchery spring chinook salmon in the Willamette basin, beginning with the 1997 brood, were marked with adipose fin clips. Although intentions were to mark all hatchery chinook, less than 100% of the returning adults will have an external mark for several reasons. First, a percentage of hatchery releases do not receive a clip because fin-

<span id="page-4-0"></span>clipping personnel do not clip the adipose fin or clip only a portion of the fin, which then regenerates. For example, about 3% of hatchery fish were released without a clip in a sample of 76 release groups from the 1996–1999 broods. Second, fry and pre-smolts without fin clips have been released in the basin. Given the large numbers of hatchery fish released, even a small percentage of unmarked hatchery fish can bias estimates of wild spawners, especially because the number of wild fish in the basin is low. To help separate hatchery fish without fin clips from wild fish, otoliths were thermally marked on all hatchery spring chinook released into the McKenzie and North Santiam rivers in the 1995 and 1996 brood years, and on all Willamette basin releases beginning with the 1997 brood year. In 2003, all returning spring chinook salmon originating from Willamette basin hatcheries should be otolith marked. Analysis of otolith marks in returning adults is scheduled to continue through the 2005 run year, which will give us three brood years (1998–2000) to evaluate the proportion of hatchery and wild fish in the unclipped portion of the run. Otolith marking may be discontinued if analyses of these brood years show that the number of unclipped hatchery fish: (1) can be predicted from the percentage of hatchery fish released without a fin clip at time of release, (2) is a minor component of the run, or (3) is a consistent proportion of the run.

#### **Methods**

#### **Juveniles**

Thermal marks were placed on otoliths of all 2002 brood, hatchery spring chinook salmon in the Willamette and Sandy basins. Reference samples were collected at the hatcheries (Table 1) and will be analyzed for mark quality at the otolith laboratory operated by Washington Department of Fish and Wildlife (WDFW). Preliminary results indicated good quality marks at all hatcheries, final results will be reported in 2004.

Table 1. Data on thermal marking of spring chinook salmon in Willamette River hatcheries and collection of reference samples, 2002 brood. Reference samples consisted of 40–50 fry (35–50 mm) from each egg take.



<sup>a</sup> *Difference between heated or chilled treatment and ambient incubation temperature.*

<sup>b</sup> Number of treatment cycles for hatched fry, except where noted.<br><sup>c</sup> 4 cycles were administered to eggs and 3 cycles to hatched fry.

#### <span id="page-5-0"></span>**Adults**

We collected otoliths from adult chinook salmon on spawning grounds and at hatcheries in most of the major tributaries in the Willamette and Sandy basins in 2003 (**APPENDIX B**). Carcass surveys were conducted throughout the spawning period to collect otoliths from spring chinook salmon without fin clips. Otoliths were removed from carcasses and placed into individually numbered vials. In addition, we collected otoliths from adult hatchery fish at Clackamas, Minto (North Santiam River), South Santiam, McKenzie, and Willamette hatcheries to serve as reference samples for blind tests of accuracy in identifying thermal marks (**APPENDIX B**). We also collected otoliths from unclipped fish at the hatcheries. Otolith samples will be sent to WDFW for analysis and will be reported in 2004.

We estimated the proportion of naturally produced ("wild") fish on spawning grounds in the Willamette and Sandy basins from otoliths collected in 2002 (Table 2). Wild fish were determined by absence of a fin clip and absence of an induced thermal mark in the otoliths. Because we saw a significant difference between the distribution of redds and the distribution of carcasses recovered among survey areas within some watersheds (Figure 1), we used the distribution of redds among survey areas to weight the number of no clip carcasses in each area. We then used results of otolith analysis to estimate the number of wild fish that would have spawned within a survey area. We reasoned that variability in counting redds among survey areas was less than that in finding and recovering carcasses because spring chinook redds are in relatively shallow water and their visibility is less dependent on stream characteristics such as stream size or survey method (boat versus foot) than that of recovering carcasses.



Table 2. Number of otoliths collected from adult spring chinook in the Willamette and Sandy basins that were analyzed for presence of thermal marks, 2002.



Figure 1. Distribution of redds and carcasses within the Clackamas and McKenzie watersheds, 2002.

<span id="page-7-0"></span>We estimated the number of wild fish in the North Santiam, McKenzie, Clackamas, and Sandy rivers above dams from the proportion of wild and hatchery fish collected in spawning surveys above the dams. The number of wild fish  $(N_w)$  was estimated using the equation:

$$
N_w = N_{\text{nc}} (1 - T_{\text{nc}})
$$

where  $N_{nc}$  is the estimated number of fish without fin clips passing over dams, and  $T_{nc}$  is the percentage of non-clipped carcasses recovered above dams with thermal marks in their otoliths.

We also estimated the number of wild fish in the McKenzie and North Santiam rivers by using the percentage of hatchery fish released without clips and the number of fin-clipped adults counted at dams to estimate the number of additional hatchery fish without a clip. Because only fin-clipped fish are harvested in fisheries, we expanded the count of fin-clipped adults at the dams by 26%, the 1981–1995 average harvest rate in the lower Willamette River sport fishery (data from Foster and Boatner 2002).

We tested the accuracy of identifying induced thermal marks by submitting otoliths to the WDFW lab from known hatchery adults as determined by adipose fin clips and coded wire tags. These samples were randomly mixed with samples collected from unclipped carcasses and were not identified as "hatchery" samples.

We used handheld tag detectors (Northwest Marine Technology, Inc.) to check for coded wire tags in carcasses with adipose fin clips. We collected the snouts of fish with a tag, which were then put into plastic bags along with an identification number.

#### **Results**

Wild spring chinook composed the highest percentage of carcasses recovered in the Sandy, Clackamas, and McKenzie rivers and the lowest percentage in the Molalla and Middle Fork Willamette rivers in 2002 (Table 3). Of interest was the high number of wild carcasses recovered in the South Santiam River. The percentage of hatchery spring chinook was relatively high in the Clackamas River above North Fork Dam despite the removal of clipped hatchery fish at the dam (Table 3). Unclipped hatchery fish with coded wire tags (double-index groups) are released in the Clackamas River and some were likely released above the dam because unclipped fish were not scanned for the presence of a coded wire tag. The percentage of no clip hatchery fish was highest in the lower reaches of the Clackamas River immediately above reservoir, particularly in the South Fork Clackamas. They were also found late in the season; 92% of all carcasses in the South Fork Clackamas were found in October compared to 47% for the rest of the Clackamas basin above the dam. Clackamas Hatchery did not take unclipped fish and they were released back into the river throughout the run. Unclipped hatchery fish migrating above the dam would have difficulty correcting their course and may have been less likely to migrate very far above the reservoir. Unclipped fish were scanned for tags at the trap in 2003.

Table 3. Composition of spring chinook salmon in the Willamette and Sandy basins based on carcasses recovered, weighted for distribution of redds among survey areas within a watershed. For comparison, the percentages of wild carcasses not weighted for redd distribution are also presented.



<sup>a</sup>*The proportion of hatchery and wild fish were determined by presence or absence of* 

*thermal marks in otoliths.* 

 $\cdot$  About 95% of the 1995 brood (5-year-old) was released without an adipose fin clip.

<sup>d</sup> *Including Fall Creek.*

<sup>e</sup>*Fish were sorted at the dams and all or most of clipped fish were removed.*

In the four rivers where we were able to estimate the number of wild spring chinook, the McKenzie River had the highest number and the North Santiam had the lowest number (Table 4). Wild and hatchery fish were more numerous in 2002 than in 2001, with a large increase of wild fish in the North Santiam River. The percentage of wild fish in the McKenzie River above Leaburg Dam decreased in 2002 (Table 4), at least in part because the number and percentage of clipped fish at Leaburg Dam increased from 20% in 2001 to 33% in 2002. In the Sandy River, an additional 157 fish without fin clips were collected at Marmot Dam on the Sandy River and were taken to Clackamas Hatchery to start a new brood stock. Of the 97 otoliths sampled from these fish, 79% were wild.

Table 4. Estimated number of wild and hatchery adult spring chinook salmon in the McKenzie, North Santiam, Clackamas, and Sandy rivers above dams. Estimated from counts at the dams and from presence of induced thermal marks in otoliths of unclipped carcasses recovered on spawning grounds. Numbers at dams were from video counts (McKenzie), daily trap counts (Clackamas and Sandy), and expanded trap counts (North Santiam, from 4 d/wk counts).



<sup>a</sup> Adjusted by distribution of redds among survey areas.<br><sup>b</sup> Escapement at Bennett Dam was likely underestimated (see Schroeder et al. 2001).

 $\degree$  Average of adjusted spawning ground samples (49.4%) and samples from Minto Pond

*(63.6%).* <sup>d</sup> *Fish were sorted at North Fork (Clackamas) and Marmot (Sandy) traps and only fish with no fin clips were allowed to pass.* 

We also estimated the number of wild fish by using the percentage of juvenile hatchery fish released without a fin clip, and compared these to estimates based on analysis of otoliths in carcasses recovered without a fin clip. In general, estimates of wild spring chinook salmon calculated from the percentage of unclipped juveniles in hatchery releases were larger than those estimated from otoliths (Table 5). Possible reasons for the discrepancy are that partially-clipped adipose fins (classified as clipped at time of release) may regenerate, or the precision in classifying adipose fins as "clipped" is greater when juvenile fish are in hand than when adults are counted on video tape or netted and passed at dams. The exception was the 2001 run in the North Santiam River, which was composed of a large number of adults with fin clips and a small number without clips. Based on juvenile release data, we estimated no wild adults after adjusting for harvest difference because of selective fisheries on fin-clipped fish. For comparison, we estimated 220 wild fish in the North Santiam in 2001 based on otoliths from carcasses without fin clips (Table 5).

Table 5. Comparison of two methods of estimating the number of wild spring chinook salmon from adult counts at dams in the McKenzie and North Santiam rivers. The proportion of wild and hatchery adults is estimated either by the percentage of juvenile hatchery fish released without fin clips or by otoliths from carcasses recovered on spawning surveys.



The WDFW otolith laboratory correctly identified a high percentage of adult hatchery spring chinook in the blind tests (Table 6). Additional tests are planned on the accuracy of identifying hatchery fish by presence of thermal marks in otoliths and identifying wild fish by absence of thermal marks.

Table 6. Accuracy of the WDFW otolith laboratory in identifying presence or absence of thermal marks in hatchery spring chinook salmon (blind tests), 2002.



The percentage of stray hatchery fish recovered in spawning surveys was much higher in the McKenzie and North Santiam rivers in 2002 (42% and 30%, respectively) than in 2001 (13% and 6%, respectively). The highest number of strays in the McKenzie and North Santiam rivers was from netpen and direct releases into the lower Willamette or Clackamas rivers (Table 7). Strays in the McKenzie River from releases into the Middle Fork Willamette and strays in the North Santiam from releases into the

Molalla were also high. The percentage of strays in other rivers ranged from 21% in the Molalla to 4% in the Middle Fork Willamette (Table 7). We collected 258 snouts from carcasses with adipose fin clips in 2003 (Table 8). Coded wire tags recovered from these fish will be read and reported in 2004.

Table 7. Origin of hatchery spring chinook salmon from recoveries of coded wire tags in spawning ground surveys, 2002. North Santiam data include recoveries in Little North Fork Santiam (2) and Middle Fork Willamette data include recoveries in Fall Creek (4).



<sup>a</sup>*McKenzie stock released in the lower Clackamas or Willamette rivers.*

b<br>B McKenzie stock reared at Willamette Hatchery and released into the lower Willamette River.<br><sup>c</sup> South Santiam stock.<br><sup>d</sup> Includes releases in Fall Creek.

<sup>e</sup> Middle Fork Willamette stock released into netpens near mouth of Columbia River.

Table 8. Number of snouts collected from carcasses of adult spring chinook salmon with adipose fin clips and a coded wire tag (determined with a hand-held detector), 2003.



<sup>a</sup> Includes 3 collected in Fall Creek.<br><sup>b</sup> Includes 5 collected below Leaburg Dam.

 $\Omega$ <sup>c</sup> 5 each collected above North Fork and Marmot dams.

### <span id="page-12-0"></span>**TASK 1.3–DISTRIBUTION AND ABUNDANCE OF NATURAL SPAWNERS**

We surveyed most of the major tributaries in the Willamette and Sandy basins in 2003 by boat and on foot to count spring chinook salmon carcasses and redds. We counted redds during peak times of spawning based on data from past surveys. In areas where we regularly surveyed to collect otoliths from carcasses, we used the highest number of redds counted in any one survey as the total number of redds for an individual section.

The North Santiam River was regularly surveyed June 27–October 15 to recover carcasses and count redds. An unusually large return of spring chinook to the North Santiam River stimulated sports fisheries and prompted reports from anglers of large numbers of pre-spawning mortalities. We began surveys in late June to confirm those reports and to examine carcasses.

In 2003 we found that 72% of females died before spawning based on recovery of carcasses in spawning surveys (Table 9). Surveys in other years began later and estimates of pre-spawning mortality may be underestimated if mortality of chinook salmon begins in early summer, as in 2003 (Table 9). However, estimates of prespawning mortality may be high if conditions such as higher flow make it more difficult to recover carcasses later in the season when most of the carcasses would be spawners. Although dead male chinook salmon were also recovered throughout the summer, they are not included in the estimate of pre-spawning mortality because later in the spawning season we cannot accurately judge if they spawned. The number of all dead salmon found in August as a proportion of the Bennett Dam counts through August was slightly higher in 2003 (3.5%) than in previous years (Table 10). Although the mean daily flow in August was lower in 2003 than in other years, the maximum water temperature was similar to 2002 and lower than 2001 (Table 10). We did not include data from 1999 and 2000 because carcass surveys were not conducted in August in 1999, and the 2000 count at Bennett was likely underestimated (*see* Schroeder et al. 2001). More detailed data are in Appendix Table C-1, and the pre-spawning mortality of other rivers is in Appendix Table C-2.

Table 9. Season total percentage (through mid to late October) of chinook salmon females that died before spawning in the North Santiam River as assessed from recovery of carcasses.



Table 10. Summary of spring chinook salmon counts through August, and number of carcasses recovered, water temperature, and flow in August in the North Santiam River, 1998, 2001–2003.



<sup>a</sup> *Mean daily maximum.*

<sup>b</sup>*Surveys began June 18, and 8 surveys were made before August 1.*

Redd digging was first observed on August 8 and peak spawning occurred in late September, similar to previous years. The redd density in 2003 was highest in the section immediately below Minto dam (55.5 redds/mi) and was almost four times that of 1996–2002 average (14.4 redds/mi, Table 11). Of the carcasses we recovered in the North Santiam in 2003, 86% had fin clips (Table 12), compared to 86% and 73% in 2001 and 2002, respectively.

Table 11. Summary of spawning surveys for spring chinook salmon in the North Santiam River, 2003, and comparison to redd densities in 1996–2002. Spawning in areas below Stayton may include some fall chinook.



<sup>a</sup>*Corrected number.*

<sup>b</sup> Data was recorded for Mehama–Stayton; density for this section was 0.9 redds/mi.<br><sup>c</sup> 400 unclipped adult spring chinook were released into the Little North Fork Santiam on August 20 and *30, September 5 and 6, 2002.* 

<sup>d</sup> *268 unclipped adult spring chinook were released into the Little North Fork Santiam in June (25th), July*  $(9^{th}, 15^{th}, 22^{nd})$ , August (25<sup>th</sup>), and September (2<sup>nd</sup>, 4<sup>th</sup>).

Table 12. Composition of naturally spawning spring chinook salmon from carcasses recovered in the North Santiam River above Stayton Island, 2003.



<sup>a</sup> Otoliths have not yet been read to determine the proportion of wild and *hatchery fish.* 

We calculated approximate fish/redd ratios for spring chinook salmon in the North Santiam basin above Bennett dams by estimating the number of potential spawners from estimates of chinook over the dams minus the number of fish removed at the Minto collection pond (e.g., fish spawned and fish transported above Detroit Dam) and those caught in the sport fishery (assuming a 20% exploitation rate). Adult chinook were transported from Minto to the Little North Fork Santiam in 2002 and 2003, and because we included redds in the Little North Fork, we did not subtract these fish from the Bennett counts. The fish/redd ratio was higher in 2003 (10.2) and 2001 (9.2) than in 2002 (6.9), which corresponds to the high pre-spawning mortality we saw in 2003 and 2001 (Table 9).

The McKenzie River was regularly surveyed August 7–October 13 to recover carcasses and count redds. Some redds were counted in August but active redd building began in early September, similar to previous years. Peak spawning occurred in late September to early October. More redds were counted in 2003 (1,187) than in 2002 (922) but relative redd densities in specific sections varied (Table 13). A large number of spring chinook were found in upper Horse and Lost creeks, areas not previously surveyed by our project. The percentage of fin-clipped carcasses in 2003 was higher above Leaburg Dam (32%) than in 2002 and 2001 (24% and 19%, respectively), but was similar below Leaburg Dam in 2003 (70%), 2002 (67%), and 2001 (72%) (Table 14). Water supply to McKenzie Hatchery was reduced in 2003 because of construction in the Leaburg Canal. Consequently, water temperature in the hatchery fish ladder was higher and likely resulted in lower effectiveness of attracting adult chinook salmon into the hatchery. McKenzie Hatchery is 3 km downstream of Leaburg Dam.

Table 13. Summary of chinook salmon spawning surveys in the McKenzie River, 2003, and comparison to redd densities (redds/mi, except redds/100 ft for spawning channel) in 1996–1998 and 2000–2002.



<sup>a</sup> *Except redds/100 ft for spawning channel.*

Table 14. Composition of naturally spawning spring chinook salmon from carcasses recovered in the McKenzie River, 2003.



<sup>a</sup> Otoliths have not yet been read to determine the proportion of wild and *hatchery fish.*

We regularly surveyed the Clackamas River basin above North Fork Dam August 20–October 20 to recover carcasses and count redds. Peak spawning generally occurred in early October, later than in previous years. A higher percentage of redds was counted above Cripple Creek in 2003 (73%) than in 1996–1999 (66%) (Table 15). Although the count of adult chinook passed at North Fork Dam was higher in 2003, our redd counts were lower and we accounted for a much lower percentage of the run (20%) than in previous years (49%) (Table 16). However, our recovery rate of chinook salmon passed at North Fork Dam (carcasses/dam count) was slightly higher in 2003 (5.3%) than in 2002 (4.7%), suggesting that pre-spawning mortality reduced the number of available spawners or that we did not count as many redds as should have been present. A higher percentage of the spring chinook run in the upper Clackamas River passed North Fork Dam in May–July in 2003 (77%) than in 1996–2002 (39%), which might increase the possibility of pre-spawning mortality in the early portion of the run. Of the female carcasses we processed above North Fork Dam, 27% died before spawning. Several factors may have affected our count of redds: (1) rain during early October discolored the main stem of the Clackamas where most redds are usually found; (2) large numbers of coho were in the upper Clackamas River (almost 1,500 passed the dam in September and early October) and were observed spawning in early October, which confounded our surveys of chinook redds; and (3) surveyors frequently encountered spawning gravels with large numbers of spawners and multiple redds in the upper Clackamas, which probably resulted in underestimated redd numbers.

The Clackamas River below River Mill Dam was surveyed July 24–October 7. We counted more redds in the upper reach of this section than in previous years (Table 15). Although fall chinook may be present below River Mill Dam, 77% of all the carcasses we processed had adipose fin clips indicating they were hatchery spring chinook. The remaining fish could be unclipped spring chinook salmon (hatchery and wild) or fall chinook. In previous years, the spring race composed 65% and 28% of the fish above and below Barton, respectively.

Table 15. Summary of spawning surveys for spring chinook salmon in the Clackamas River basin, 2003, and comparison to redd densities in 1996–1999 and 2002.



<sup>a</sup> Additional 33 carcasses were processed in the 0.3 mi River Mill Dam–McIver section. b<sup>b</sup> Scales have not been read to separate spring chinook salmon from fall chinook *salmon.*

 Table 16. Counts of adult spring chinook salmon above North Fork Dam and the relationship to successful spawners above the dam, 1996–1999, 2002–2003.



<sup>a</sup> *Total from video counts (1996–1998) or fishway trap counts (after 1999) up to one week prior to last spawning survey.* <sup>b</sup> *Estimated from redds using 1:1 sex ratio and two fish per redd.* <sup>c</sup> *Fish from dam count divided by redds*.

We regularly surveyed the Sandy River basin above Marmot Dam August 19– October 15 to recover carcasses and count redds (Table 17). Peak spawning generally occurred in late September to early October, similar to other years. Distribution of redds was similar to 2002, in that a lower percentage of redds in the basin was counted in the Salmon River in 2003 (68%) than in 1996–1999 (81%), and a higher percentage of redds was counted in Still Creek and the Zigzag River in 2003 (28%) than in previous years (16%). We accounted for a lower percentage of the spring chinook salmon run over Marmot Dam in 2003 (37%) than in 2002 (47%), and it was similar to that in 1999 (34%) (Table 18).

We surveyed the Sandy River below Marmot Dam in 2003 on September 24–25, and counted 183 redds (Table 17). Most of the chinook salmon found below Cedar Creek were not clipped (70%), whereas just 25% of the fish recovered between Cedar Creek and Marmot Dam were not clipped. Sandy Hatchery is located on Cedar Creek.



Table 17. Summary of spawning surveys for spring chinook salmon in the Sandy River basin, 2003, and comparisons to redd densities in 1996–1999, 2002.

<sup>a</sup> *Survey did not extend to the mouth as in other years.*

<sup>b</sup>*82 additional carcasses not processed.*



<span id="page-19-0"></span>Table 18. Counts of adult spring chinook salmon at Marmot Dam and the relationship to successful spawners in the Sandy River basin above the dam, 1996–1999, 2002–2003.

<sup>a</sup> *Total from video counts (1996–1998) or fishway trap counts (1999, 2002) up to one* 

week prior to last spawning survey.<br>
<sup>b</sup> Sandy River above dam from punchcard estimates. No fishery after 1998.<br>
<sup>c</sup> Estimated from redds using 1:1 sex ratio and two fish per redd.<br>
<sup>d</sup> Fish from dam count minus harvest di

Other rivers that were regularly surveyed in 2003 (Table 19) were South Santiam (7 dates, 14 July–21 October), Molalla (4 dates, 27 August–7 October), and Middle Fork Willamette (6 dates, 15 July–29 September). Active redd building began in early September in the South Santiam and in mid September in the Middle Fork Willamette. Peak spawning in both rivers was late September to early October. The percentage of fin-clipped carcasses was lower in the Middle Fork Willamette (54%) than in the North Santiam (86%), South Santiam (84%), and Molalla (79%) (Tables 12 and 19).

#### **TASK 2.1– MORTALITY IN A CATCH AND RELEASE FISHERY**

We conducted a study of hooking mortality of spring chinook salmon in the lower Willamette River sport fishery in 1998–2000. A manuscript describing this study was submitted and accepted for publication (Lindsay et al. in press). The paper is scheduled to be published in May 2004.

<span id="page-20-0"></span>



<sup>a</sup>*Otoliths have not yet been read to determine the proportion of wild and hatchery fish.* 

#### **TASK 3.1– EVALUATION OF NET PENS IN THE LOWER WILLAMETTE RIVER**

In the 1970's, studies by Smith et al. (1985) found that trucking juvenile spring chinook salmon below Willamette Falls at Oregon City increased angler catch in the Clackamas and lower Willamette rivers by improving survival to adult return. Straying also increased. However, Specker and Schreck (1980) found that trucking smolts caused severe stress that tended to reduce survival compared to fish not trucked. Johnson et al. (1990) and Seiler (1989) suggested that stress from trucking could be reduced and survival increased by acclimating juveniles at a site for several weeks before release. Acclimation at lower river sites may increase angler harvest by improving survival of juveniles and by delaying migration to upriver areas.

A study was begun in 1994 to determine if acclimation prior to release could be used to increase sport harvest of hatchery spring chinook salmon returning to the lower Willamette River. We used McKenzie River stock in the study because of concerns about straying of other stocks into the McKenzie, a stronghold for wild spring chinook salmon. The evaluation of straying was an important part of the study. Fish were acclimated in net pens and compared to fish trucked directly from the hatchery. Control groups were released into the McKenzie River from McKenzie Hatchery. The study was <span id="page-21-0"></span>originally planned for four brood years. However, numerous problems led to modifications in study design beginning with the 1995 brood and an extension of the study for four additional years through 1999 brood releases. Smolt releases from 1992– 1999 broods are described in Lindsay et al. (1997), Lindsay et al. (1998, 2000), and Schroeder et al. (1999, 2001). The types of experimental groups released in all brood years are summarized in Schroeder et al. 2002.

#### **Adult Recapture of 1995–1996 Brood Releases**

Coded wire tags from experimental releases were primarily recovered from adults captured in fisheries, in hatcheries, in traps at dams and on spawning grounds. Most of the sport fishery for spring chinook salmon in the Willamette River occurs below Willamette Falls. Although some catch of spring chinook salmon occurs above Willamette Falls, these fisheries generally are not surveyed. Based on salmon catch card records, the fishery above Willamette Falls accounted for about 26% of the total basin harvest annually in 1981–1995 (calculated from Foster and Boatner 2002). Adult captures from 1992 through 1994 broods and conclusions based on these data are reported in Schroeder et al. 2002.

Adult captures from 1995 and 1996 broods are shown in Tables 20 and 21. The 1996 brood represents the first of four consecutive brood years with duplicated releases, which should help identify differences among groups after all four broods have returned. For 1995 and 1996 brood tag recoveries to date, two tentative conclusions can be reached. First, the conclusion in Schroeder et al. 2002 that smolt releases into the lower Willamette River do not increase sport catch appears to be holding true through 1996 brood recaptures. Sport catch below the falls of control fish released from McKenzie Hatchery was equal to or higher than catch of fish from groups acclimated or released directly into the lower main stem Willamette (Multnomah Channel for 1995 and 1996 broods). Second, for groups released into the Clackamas River in spring, those acclimated in Clackamas Cove appear to contribute more to sport fisheries in the Willamette and Clackamas rivers than groups released directly into the Cove or into the Clackamas River. However, for the 1995 brood, returns from acclimated groups in the Cove contributed less to the sport fishery than did the control.

Table 20. Capture of adult spring chinook salmon from the net pen evaluation of smolt releases into the lower Willamette River basin, 1995 brood. Numbers were adjusted to a standard release of 100,000 smolts. Data were obtained from the coded wire tag database of the Pacific States Marine Fisheries Commission, November 2003.



Table 21. Capture of adult spring chinook salmon from the net pen evaluation of smolt releases into the lower Willamette River basin, 1996 brood. Numbers were adjusted to a standard release of 100,000 smolts. Data were obtained from the coded wire tag database of the Pacific States Marine Fisheries Commission, November 2003. Data are preliminary.



<sup>a</sup> Counts are underestimated because adult returns have not yet been adjusted for sub-sampling at hatcheries in 2001.

#### <span id="page-24-0"></span>**TASK 3.4– INCORPORATING WILD FISH INTO HATCHERY BROODSTOCKS**

Otoliths were collected in 2002 from spring chinook salmon without fin clips that were spawned at Willamette basin hatcheries to determine the number of wild fish that are being incorporated in the broodstocks. The highest percentage of wild fish in the unclipped portion of the broodstock was in South Santiam Hatchery, which also had the highest percentage of wild fish incorporated into their broodstock (Table 22).

Table 22. Composition of spring chinook salmon without fin clips that were spawned at Willamette basin hatcheries, based on the presence or absence of thermal marks in otoliths, 2002.



#### **TASK 4.1– MIGRATION TIMING OF WILD JUVENILE SPRING CHINOOK SALMON**

We started field work in 1999–2000 under Objective 4 of our project study plan (**APPENDIX A**). Information collected under Objective 4 will allow managers to better understand spatial and temporal use of habitat by juvenile wild spring chinook in the Willamette basin and to better protect existing natural production areas. We initially began work on wild chinook in the McKenzie River where three life history types were defined at Leaburg Dam: (1) age 0 fry that migrate in late winter through early spring, (2) age 0 fingerlings that migrate in fall, and (3) yearling smolts that migrate in early spring. Initial work concentrated on determining juvenile migration timing of these three life history stages below Leaburg Dam in the McKenzie and Willamette rivers. In 2002 and 2003, our work expanded into the lower Willamette River and in the Santiam River basin where juvenile fall chinook salmon may be present.

In an effort to better define the distribution and migration timing of wild spring chinook, we began work in cooperation with NOAA Fisheries (Conservation Biology Division, Northwest Fisheries Science Center, Seattle) on use of DNA microsatellite analysis to identify the two races of chinook salmon. Fall chinook salmon were introduced above Willamette Falls in 1964 and about 5–12 million smolts were released each year until 1996 when the hatchery program was discontinued. Most fall chinook returned as age 3 (69%) or age 4 (18%) during the late 1990s (Foster and Boatner 2002). The average count at Willamette Falls in 1990–2000 (when the last age 4 fish

<span id="page-25-0"></span>from hatchery releases would have returned) was about 4,100 adults (Foster and Boatner 2002). The 2001–2003 count averaged 955 adults, with just 390 fish returning in 2003. In recent years, most of the fall chinook salmon have spawned in the lower Santiam River.

#### **Methods**

We used PIT tags (Prentice et al. 1990a, 1990b) to monitor migration of juvenile spring chinook salmon in the McKenzie and Willamette rivers. We injected fish with 134.2 kHz tags, and used a tag detector (Destron-Fearing® FS2001F), a laptop computer, and a computer program developed by Pacific States Marine Fisheries Commission (PSMFC) to enter data. All tagging data were loaded into a PIT tag database (PTAGIS) maintained by PSMFC.

Age 0 juvenile chinook salmon migrating in fall 2002 and yearling smolts migrating in spring 2003 were collected with a rotary screw trap installed in the Leaburg Dam bypass flume. Age 0 chinook salmon representative of the fry migrants were seined and tagged in the lower McKenzie and upper Willamette rivers in June and July because fry are too small to tag when they migrate past Leaburg Dam in February– April. We confined our sampling to the lower McKenzie and upper Willamette rivers downstream of spawning reaches to insure the juvenile chinook salmon we tagged had migrated. We also tagged a sample of hatchery fish that were released in the fall and spring from McKenzie Hatchery. In addition, we seined in sections of the Willamette River from Harrisburg to Newburg and in the Santiam River basin.

Migrating juvenile chinook salmon were scanned with a tag detector (Destron-Fearing® FS1001) at Willamette Falls in the bypass system of the Sullivan hydroelectric plant operated by Portland General Electric Company (PGE). Only a portion of the juvenile salmon migrating past Willamette Falls uses the bypass system (Royer et al. 2001). Additional tags were detected and reported by the National Marine Fisheries Service during their juvenile salmonid studies in the Columbia River estuary (Ledgerwood et al. 2000).

Because most of our tags were detected by passive interrogation, we did not measure growth between time of tagging and time of detection. We used fork lengths (FL) of individual fish at the time they were tagged to examine differences between the mean length of all tagged fish and the mean length of detected fish. We compared the difference in mean length by considering all tagged fish as a known population (N) and the detected fish as a sample (n) of N. We statistically analyzed differences in mean length by calculating a *t* value using the following equation:

$$
\frac{(\bar{FL}_N - \bar{FL}_n)}{\sqrt{v_n(1-p)/n}}
$$

<span id="page-26-0"></span>where  $\overline{FL}_N$  is the mean length of all juvenile fish given PIT tags,  $\overline{FL}_n$  is the mean length at the time of tagging of fish that were later detected,  $v<sub>n</sub>$  is the variance of the mean length of detected fish, and p is the detection rate of tagged fish (Willamette Falls and Columbia River). The square root of 1-p is the finite population correction (Snedecor and Cochran 1980).

Initial genetic analyses were conducted to investigate the feasibility of using DNA microsatellites to separate spring chinook and fall chinook. Samples in the initial analyses were tissues from frozen juvenile spring chinook that had been previously collected in the Clackamas, North Santiam, and McKenzie rivers for electrophoretic studies (Lindsay et al. 1997), and tissues from adult fall chinook collected at Willamette Falls in September 2002. We submitted additional reference samples in 2003 that included tissues collected in 2002 of adult hatchery spring chinook from Marion Forks (48) and McKenzie (48) hatcheries, and of wild spring chinook from spawning areas in the upper North Santiam (27) and McKenzie (48) rivers.

Tissues were collected from a sample of juvenile chinook captured by beach seine in the summer of 2002 (Schroeder et al. 2002) in anticipation that they could be identified by race using DNA microsatellite analyses. We collected tissues June 19–24 from juvenile chinook salmon in the Willamette River above and below the mouth of the Santiam River (15 and 64 samples, respectively), and in the lower Santiam River (71). We also collected tissues July 30–31 in the Willamette River below the Santiam River (37). These samples were analyzed by NOAA Fisheries.

#### **Results**

We tagged 6,578 wild spring chinook salmon in the McKenzie and Willamette rivers, 1,991 hatchery fish from McKenzie Hatchery, and 712 wild chinook salmon in the lower Willamette and Santiam rivers in June 2002–April 2003 (Table 23). In addition, we tagged over 3,800 age 0 wild chinook salmon in the lower McKenzie and upper Willamette rivers, and over 2,200 in the lower Willamette River and Santiam basin in spring and summer 2003 (Table 24).

Most of the detections of fish tagged in June 2002–April 2003 occurred at Willamette Falls (Table 23). Detection rates at Willamette Falls were generally low in 2002–2003 (Table 25). The efficiency of the passive interrogator depends on river flow, which affects the proportion of juveniles using the bypass system at the Sullivan Plant and the amount of debris in the bypass system. In high flows, fewer fish migrate through the bypass system and the interrogator is more difficult to operate because of debris. In previous years, flows in November–January appeared to have the largest influence on the efficiency of the interrogator. However, flows in March appeared to have the greatest influence on tag recoveries this year. The average flow in March 2003 was 150% above the 1965–2002 average, and the interrogation system was fully operational less than 50% of the month (Appendix Table D-4). As an example, shortly after the first tagged hatchery fish from the spring release passed the falls on March 8,



Table 23. Detection of juvenile wild and hatchery spring chinook salmon given PIT tags and released in June 2002–April 2003. Tags were detected at the PGE Sullivan Plant at Willamette Falls unless noted.

<sup>a</sup> Sullivan Plant tag detector operated October 1, 2002 – September 30, 2003 with occasional shutdowns.<br><sup>b</sup> Includes two fish detected in Columbia River estuary (rkm 76).

the interrogator was shut down for several days and the flow increased to 64,000 cfs. Flows remained high through most of March and April, which has been a period of high migration in other years for chinook tagged in the previous summer and fall at age 0. An automatic brush-cleaning system was installed by PGE on the dewatering screen panels in summer 2003 to help keep the interrogator operational during periods of high debris.

Table 24. Number and mean fork length of wild spring chinook salmon (age 0) that were seined, PIT-tagged, and released in the McKenzie River below Hendricks Bridge (rkm 34), in the Willamette River above and below the Santiam River, and in the Santiam River watershed, 2000 (July–September), 2001 (July–August), 2002 (June– July), and 2003 (late May–mid July).



<sup>a</sup> An additional 67 and 136 hatchery fish were seined and tagged in the McKenzie and Upper Willamette rivers, respectively.

<sup>b</sup> Does not include four hatchery fish *Does not include four hatchery fish Contingent* **by** *From confluence of North and South Santiam to mouth.* 

Table 25. Detection rate (%) at Willamette Falls of spring chinook salmon that were PIT-tagged and released in Willamette River basin in October 1999–April 2003.



Almost all the wild chinook salmon tagged as age 0 fish in summer 2002 migrated in the summer and fall, whereas all the wild fish tagged in fall 2002 migrated the following spring (Figure 2), similar to the migration of age 0 fish tagged in 2001 (Schroeder et al. 2002). In contrast, all age 0 fish tagged in 2000 migrated primarily in the spring (Schroeder et al. 2001). The migration of hatchery fish peaked at Willamette Falls shortly after their release (Figure 2), similar to previous years. The peak migration at Willamette Falls of chinook salmon smolts tagged in the spring was in April and May. The migration time for fish tagged in October 1999–April 2003 is presented in Table 26.



Figure 2. Migration timing of juvenile spring chinook salmon past Willamette Falls, 2002–2003. Based on detection of fish given PIT tags in the McKenzie, Willamette, and Santiam rivers. The number of detected tags is given in parentheses in the legend.

Table 26. Travel time (median days) to Willamette Falls of juvenile chinook salmon tagged and released in the Willamette River basin in October 1999–April 2003. Number of tag detections is in parentheses.



The mean fork length of juvenile chinook salmon tagged in 2002–2003 and later detected was generally larger than the mean fork length of all fish tagged and released (Table 23), but the difference was significant (*P* < 0.05) only for wild fish released in summer and for hatchery fish released in fall.

Age 0 chinook salmon were found throughout the lower McKenzie, upper and lower Willamette, and lower Santiam rivers. The catch of juvenile chinook salmon was higher in 2003 than in previous years (Table 27), in part, because we began sampling earlier in the year before some of the fish had migrated out of the Willamette and Santiam rivers. We documented an early summer migration of age 0 chinook salmon past Willamette Falls that were tagged in all areas of the Willamette, McKenzie, and Santiam rivers (Table 28). The migration of juvenile chinook was highest for fish tagged in the Santiam basin (Table 28), possibly because these are composed of a higher number of fall chinook than in other areas (*see* genetics results below). The mean fork length of the tagged juvenile chinook salmon that were later detected was larger than the mean fork length of all the fish that were tagged (Figure 3).



Table 27. Catch rate with a beach seine (fish/seine set) of juvenile chinook salmon in the Willamette, McKenzie, and Santiam rivers, 2000–2003.

Table 28. Detection rate (%) and travel time (median days) of age 0 juvenile chinook salmon that were PIT-tagged and released in Willamette River basin in late May and June 2003, and detected at Willamette Falls in late May–late July 2003.





Figure 3. Comparison of the mean fork length (at time of tagging) of all age 0 juvenile chinook salmon that were PIT-tagged and released in Willamette River basin in late May and June 2003, and of those that were detected at Willamette Falls in late May– late July 2003. Asterisks indicate significant differences (*P* < 0.01).

The mean length of spring chinook salmon in the McKenzie and Willamette rivers tended to increase in 2003 from spring to early summer for fish sampled in similar areas (Figure 4). Comparisons of length between years generally showed that juvenile spring chinook were larger in 2000 and 2001 than in 2002 and 2003 (Figure 5).



Figure 4. Mean fork length  $(+$  SD) of juvenile chinook salmon that were seined in the McKenzie and Willamette rivers, 2003. Columns with different letters within areas are significantly (*P* < 0.05) different.



Figure 5. Mean fork length  $(+$  SD) of juvenile chinook salmon that were seined in similar locations and times of year in the McKenzie and Willamette rivers, 2000–2003. Columns with different letters within areas and times of year are significantly (*P* < 0.05) different; numbers above the bars are sample sizes.

Genetic analysis of tissue samples collected from chinook salmon of known stock origin in the Willamette basin showed distinct population groupings and demonstrated the feasibility of racial classification (Teel et al. 2002). Of 3,000 individuals that were simulated from allele frequency data, 96% were correctly classified into fall or spring chinook classifications.

Analyses of juvenile salmon of unknown run assigned most fish to spring or fall run with probabilities > 0.90 (Teel et al. 2002). About 14% of the unknown juveniles had assignment probabilities of about 0.50, which was likely because they were from populations that were insufficiently represented in the reference collections. All juvenile chinook salmon sampled in the Willamette above the Santiam River were spring chinook (Table 29). Of the 101 fish sampled in the Willamette River below the Santiam River, 94% were spring chinook. The percentage of spring chinook salmon in the lower Santiam River (66%) was lower than in the lower Willamette River (Table 29). Although the number of fall chinook counted at Willamette Falls has been low in recent years, most spawn in the lower Santiam River.



Table 29. Racial classification by DNA microsatellite analysis of juvenile chinook salmon collected by beach seine in the Willamette and Santiam rivers, 2002.

<sup>a</sup> The genotype was unable to be determined for 2 samples collected in June in the *lower Willamette and for 1 sample in the Santiam.*

Schools of juvenile chinook salmon  $(≥10$  fish) sampled at five sites below the Santiam River were mixtures of spring and fall chinook. The average proportion of spring chinook was the same (82%) in three schools sampled in June and in two schools sampled in July. The mean fork length of spring chinook and fall chinook salmon was not significantly different ( $P > 0.05$ ) in the June or July samples (Table 30).

Table 30. Mean fork length (FL [mm]) of juvenile spring and fall chinook salmon captured in the Willamette River, 2002. Race was identified by DNA analysis.



Additional juvenile tissue samples were collected in summer 2003 in the Willamette River downstream of the Santiam River (Table 31). These samples will be analyzed in 2004 by NOAA Fisheries.

Table 31. Number of tissue samples collected from juvenile chinook salmon captured by beach seine or trap in the Willamette River below the Santiam River, 2003.



### **TASK 5.3–EFFORTS TO RE-ESTABLISH POPULATIONS**

<span id="page-35-0"></span>We reported the poor survival in 2002 of 400 unclipped adult spring chinook that were transported from the Minto collection facility on the North Santiam River and released into the Little North Fork Santiam (Schroeder et al. 2002). Few of these fish survived to spawn and the number of redds counted in 2002 (30) was only slightly higher than the 1997–2001 average (20). In 2003, we increased our efforts to monitor the outplantings of adult chinook salmon from some hatcheries.

Unclipped adult spring chinook, collected at Minto, were tagged with uniquely numbered Floy<sup>®</sup> tags and released at two locations in the Little North Fork Santiam River: Golf bridge (rkm 20) and Elkhorn bridge (rkm 27). A total of 268 fish were released on six dates from June 25 through September 4 (Table 32).

We examined 46 carcasses for fin clips and tags in nine surveys from July 10 to October 6, and collected otoliths and scales from unclipped fish. An additional 25 fish were decayed and we were unable to determine if they were clipped or tagged. The first spawned female was found on September 19. Of the 268 tagged fish released in the river, we recovered 32 tags and found 10 carcasses with a tag wound. Of the 15 tagged females recovered, 14 died prior to spawning. Half of the carcasses from the Elkhorn bridge releases were recovered in the same area and half were recovered downstream. Most of the carcasses from the Golf bridge releases were recovered in the same area (75%) and the rest were recovered upstream.



Table 32. Summary of adult chinook salmon released and recovered in the Little North Fork Santiam, 2003.

 $\frac{a}{a}$  10 additional fish were recovered with tag wounds but no tag.

We counted 31 redds in the Little North Fork between Elkhorn bridge and the mouth, far fewer than expected from a release of 268 adults. However, recoveries of tagged female carcasses suggest that pre-spawning mortality was high (93%). In addition, only 8 of the 42 tagged carcasses we recovered were found after September

<span id="page-36-0"></span>19, the date of first redd deposition, which suggests that only about 19% of the 268 adults (51 fish) might have survived to spawn.

On August 29, 135 adult chinook salmon with fin clips were transported from South Santiam Hatchery and released into the Calapooia River above Bigs Cr (rkm 93). Live chinook salmon counted at the release site decreased from over 100 fish on August 30 to 6 fish on September 12 (Table 33). The cumulative number of carcasses recovered in the river (3000 line bridge, rkm 95, to Bigs Cr) increased from 2 on August 30 to 49 on September 12. We collected 43 clipped chinook salmon, 5 unclipped fish, and 11 fish that were too decayed to process. The first redd was observed on September 25 in the section from McKinley Creek (rkm 98) to the 3000 line bridge, and we counted two redds through October 8 in this section, the only redds found in 7.6 mi we surveyed above Bigs Creek.

Table 33. Observations of chinook salmon in the Calapooia River, July–October 2003.



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# **APPENDIX A**

<span id="page-40-0"></span>**Schematic of Willamette Spring Chinook Salmon Study Plan** 



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# **APPENDIX B**

<span id="page-42-0"></span>

# **Otoliths Collected from Adult Spring Chinook Salmon, 2003**

# Appendix B. Continued.



## **APPENDIX C**

# <span id="page-44-0"></span>**Data on Pre-Spawning Mortality in the Willamette and Sandy Basins, 2001–2003**

Appendix Table C-1. Summary of chinook salmon counts, pre-spawning mortalities, maximum water temperatures, and flows in the North Santiam River by two week periods, 2001–2003.



<sup>a</sup> Carcass sampling did not start until August 14 so this number underestimates the mortality in the first two weeks of August.



Appendix Table C-2. Number and percentage of carcasses of spring chinook salmon (females) in the Willamette and Sandy River basins that died before spawning and starting dates of spawning surveys, 2001–2003.

## **APPENDIX D**

#### <span id="page-46-0"></span>**Migration and Rearing Data for McKenzie, Willamette, Calapooia, and Santiam Rivers, 2002**

Appendix Table D-1. Fish species and numbers caught in seines in the McKenzie River (rkm 0–34), May 21–August 1, 2003





Appendix Table D-2. Fish species and numbers caught in seines in the upper (rkm 173–282) and lower Willamette (rkm 88–173) River, May 22–July 28, 2003.

Appendix Table D-3. Fish species and numbers caught in seines in the Calapooia (rkm 1), Santiam (rkm 16), North Santiam (rkm 1–55), and South Santiam (rkm 5–53) rivers, May 22–June 24, 2003.





Appendix Table D-4. Dates the PIT tag interrogator in the PGE Sullivan Plant at Willamette Falls was operational September 2002–September 2003.